



**PLECKSTRIN HOMOLOGY AND GREEN FLUORESCENT FUSION  
PROTEIN IN STARFISH BY USING BIOINFORMATICS**

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**Abstract**

Different signaling and metabolic processes, including those that occur during fertilization, are tightly controlled by PLC isozymes. In this research, Bioinformatic databases will be utilized to fuse PLC's PH domain with GFP, which will then be used to study the starfish *Patiria miniata*'s starfish chromosomes.

**Keywords:** Infertility, Phospholipase C, GFP, *Patiria miniata*

**Introduction**

In order to stimulate embryogenesis, cytoplasmic free  $Ca^{2+}$  is detected in the eggs. increase at fertilization [1] [2]. Anti-vertebrate protein inhibitors have been used in research in starfish eggs have been offered that this  $Ca^{2+}$  rise necessitates SFK (Src family kinase) in the egg that either activates PLC-gamma directly or indirectly, resulting in  $IP_3$  production, it causes  $Ca^{2+}$  to be released from the endoplasmic reticulum of the egg (ER). *Asterina miniata* PLC-gamma was obtained from oocyte cDNA to study the endogenous measures in starfish eggs necessary for  $Ca^{2+}$  release during fertilization in greater detail. AmPLC gamma is a cDNA that encodes a protein that is 49 percent identical PLC-gamma in mammals. Recombinant Src homology 2 (SH2) domains in AmPLC-gamma interacted with a 58-kDa Src family kinase in a fertilization-responsive way [3] [4]. PLC from a sea urchin egg immunoprecipitates the PLC-gamma was shown to be phosphorylated in response to fertilization when it was tested with an antibody specific against AmPLC-gamma. Adding starfish eggs to the mix with AmPLC tandem gamma's SH2 domains (which block activation of PLC gamma) prevented release of  $Ca^{2+}$  at fertilization. These findings show that an endogenous starfish egg PLC-gamma interacts with an egg SFK and, via a PLC-gamma SH2-mediated mechanism, mediates  $Ca^{2+}$  release during fertilization [5] [6]. Calcium signaling levels are maintained by the isoform  $PLC_{\gamma}$ , which assist to open a channel that allows for  $Ca^{2+}$  infusion over the plasma membrane and out of the endoplasmic reticulum, respectively [7]. PLC1 and PLC2 are two isoforms of the PLC class, growth factor stimulation of receptor and non-receptor (cytosolic) protein tyrosine kinase activation by polypeptide growth factor resulting in an increase in the activity of phospholipase, which can lead to angiogenesis, cell motility,



ventricular contractility, among other things [8] [9]. GFP (Green fluorescent protein) is a widely recognized and transiently expressed fluorescent tag that can play a vital function in the localization of PLC $\gamma$ . GFP will be fused to PLC's PH-domain. In this work, and the PH-GFP fusion protein will be utilized to investigate localisation in the starfish *Patiria miniata* [10]. Additional PLC family members have been demonstrated to influence Ca<sup>2+</sup> signaling via previously undiscovered mechanisms, which suggests that this fusion protein may also Other non-membrane cytosolic proteins interact with and localize to compartments when exposed to sperm-egg interaction. Because the PH domain aids in marked protein-protein, protein-lipid interactions, and membrane binding it is used in this work for the development of fusion proteins that can bind to membranes [11] [12].

## Materials and Methods

1-NCBI The NCBI houses a series of (computer files full of information) clearly connected with or related to (science that uses living things to improve the Earth) and natural communitydicine and is an important useful thing supply for bioinformatics tools and services [13]. Major computer files full of information include GenBank for DNA sequences and PubMed, a related to a list of references, computer file full of information for the study of how life and medicine work together. Other computer files full of information include the NCBI Epigenomics. All these computer files full of information are available online through the Entrez search engine [14].

2- Bioinformatics To construct a PH-GFP fusion protein, PLC PH domain of starfish PLC was amplified using bioinformatics to construct primers containing BsrG1 restriction sites the PJV53 – PAGFP plasmid [15] [16]. The NCBI database was used to retrieve the cDNA sequence in its entirety and the PH domain of AmPLC $\gamma$ . PH domain was amplified by using NCBI's Primer-Blast to build the forward and reverse primers [17] [18] [19].

## Result and Discussion

### Asterina miniata phospholipase C-gamma mRNA, complete cds

GenBank: AY486068.1

[FASTA](#) [Graphics](#)

Go to:

```
LOCUS       AY486068                3816 bp      mRNA      linear     INV 14-APR-2004
DEFINITION Asterina miniata phospholipase C-gamma mRNA, complete cds.
ACCESSION  AY486068
VERSION    AY486068.1
KEYWORDS   .
SOURCE     Patiria miniata (bat star)
  ORGANISM Patiria miniata
            Eukaryota; Metazoa; Echinodermata; Eleutherozoa; Asterozoa;
            Asterozoa; Valvatacea; Valvatida; Asterinidae; Patiria.
REFERENCE  1 (bases 1 to 3816)
AUTHORS   Runft,L.L., Carroll,D.J., Gillett,J., Giusti,A.F., O'Neill,F.J. and
            Folts,K.R.
TITLE     Identification of a starfish egg PLC-gamma that regulates Ca2+
            release at fertilization
JOURNAL    Dev. Biol. 269 (1), 220-236 (2004)
PUBMED    15081369
REFERENCE  2 (bases 1 to 3816)
AUTHORS   Gillett,J., Carroll,D.J., Runft,L.L., O'Neill,F.J., Giusti,A.F.,
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Figure 1. Initial results page of the nucleotide search for *Asterina miniata* phospholipase C-gamma mRNA. [https://www.ncbi.nlm.nih.gov/nucleotide/40365362?log\\$=activity](https://www.ncbi.nlm.nih.gov/nucleotide/40365362?log$=activity)



## phospholipase C-gamma [Patiria miniata]

GenBank: AAR85355.1

[Identical Proteins](#) [FASTA](#) [Graphics](#)

Go to:

LOCUS AAR85355 1261 aa linear INV 14-APR-2004  
 DEFINITION phospholipase C-gamma [Patiria miniata].  
 ACCESSION AAR85355  
 VERSION AAR85355.1  
 DBSOURCE accession [AY486068.1](#)  
 KEYWORDS .  
 SOURCE Patiria miniata (bat star)  
 ORGANISM [Patiria miniata](#)  
 Eukaryota; Metazoa; Echinodermata; Eleutherozoa; Asterozoa;  
 Asteroidea; Valvatacea; Valvatida; Asterinidae; Patiria.  
 REFERENCE 1 (residues 1 to 1261)  
 AUTHORS Runft,L.L., Carroll,D.J., Gillett,J., Giusti,A.F., O'Neill,F.J. and  
 Foltz,K.R.  
 TITLE Identification of a starfish egg PLC-gamma that regulates Ca<sup>2+</sup>  
 release at fertilization  
 JOURNAL Dev. Biol. 269 (1), 220-236 (2004)  
 PUBMED [15081369](#)  
 REFERENCE 2 (residues 1 to 1261)  
 AUTHORS Gillett,J., Carroll,D.J., Runft,L.L., O'Neill,F.J., Giusti,A.F.,  
 Jaffe,L.A. and Foltz,K.R.  
 TITLE Direct Submission  
 JOURNAL Submitted (24-NOV-2003) Biological Sciences, Florida Tech, 150 West  
 University Blvd., Melbourne, FL 32901, USA

ORIGIN

```

.      1 matnslykkk ltpqevasvt kmkmgvtlt rfygkrrper rsfeicmetr qilwrrqtgr
      61 tdgavkirei keirpgknsr dferwpdeak kydtslclvi cygaefrlks lsvvagnade
     121 rhkwivglnw lvedhkissy psrlewllrr efyamgktn dtvsldrmdks fmpyvnlknn
     181 tkdlkeyfne vdrwnkqeig fdgfvqlyhn lifqrevadr fkeyidernl vtvngmirfl
     241 aeqqkdttan npiavkamme sfltdlgrpc qesdpkftvp efllylfspd neiwdkkkfd
  
```

Figure 2. The Asterina miniata src cDNA was translated to Patiria miniata src protein. Amino acid sequence of the Patiria miniata Src family kinase protein, with the ph domain highlighted in brown from range( 23-141).

<https://www.ncbi.nlm.nih.gov/protein/40365363>

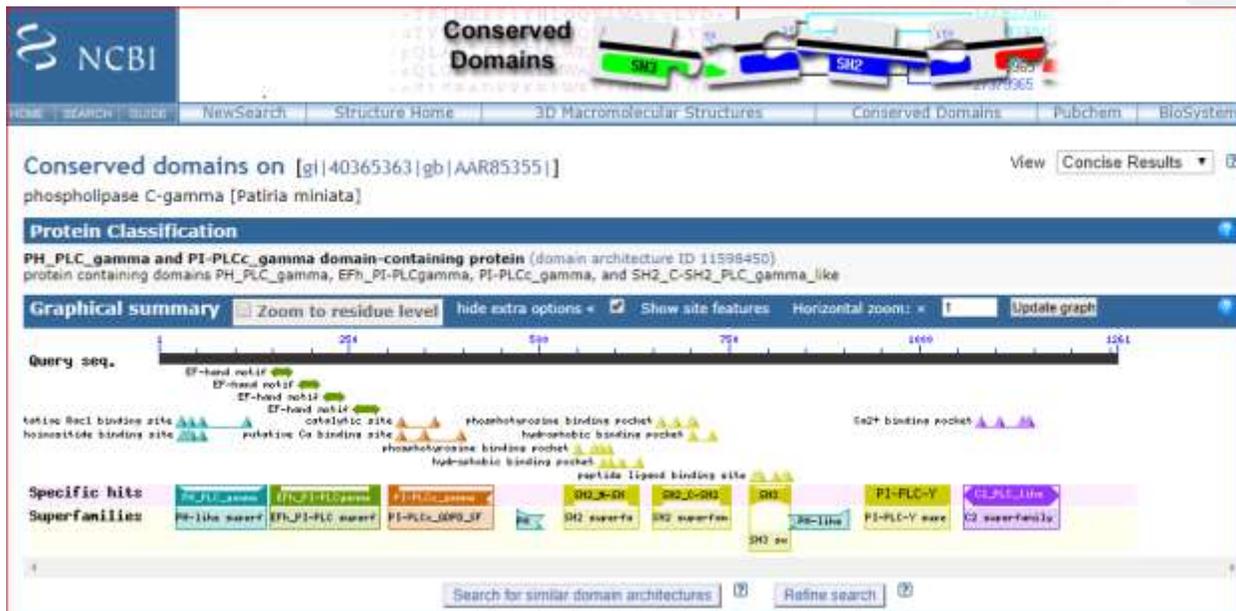
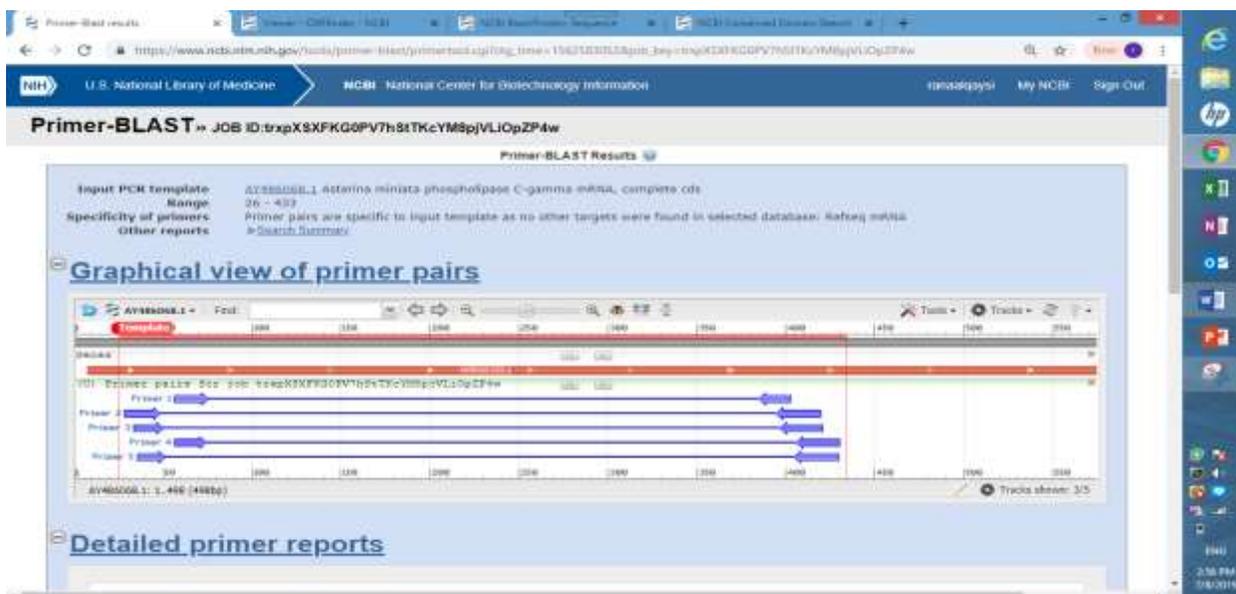


Figure 3. Conserved domain results for *patiria minita*. Conserved domain compares this protein sequence to the sequence for the same protein in other animals and identifies regions of high similarity (conserved regions). This shows that the ph domain is highly conserved in this protein. [https://www.ncbi.nlm.nih.gov/Structure/cdd/wrpsb.cgi?INPUT\\_TYPE=live&SEQUENCE=AAR8535](https://www.ncbi.nlm.nih.gov/Structure/cdd/wrpsb.cgi?INPUT_TYPE=live&SEQUENCE=AAR8535)  
5.1





Primer pair 2									
	Sequence (5'→3')	Template strand	Length	Start	Stop	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	AAGAAGAAGCTGAGGCCCA	Plus	20	29	48	61.77	55.00	4.00	2.00
Reverse primer	AGATTTATGGTCTCGACTAGCCA	Minus	25	419	395	59.81	40.00	4.00	2.00
Product length	391								

[https://www.ncbi.nlm.nih.gov/tools/primer-blast/primertool.cgi?ctg\\_time=1562583052&job\\_key=trxpXSXFKGoPV7hStTKcYM8pjVLiOpZP4w](https://www.ncbi.nlm.nih.gov/tools/primer-blast/primertool.cgi?ctg_time=1562583052&job_key=trxpXSXFKGoPV7hStTKcYM8pjVLiOpZP4w)  
Figure 4: primer-blast from NCBI are used to design forward primer and reverse primer , Next these primers will cut out by using restriction enzyme BrsGI

ORIGIN	
1	cttcagaatg gccaccaaca gcctctacaa gaagaagctg acgccccagg aggtggccag
61	cgccaccaag atgctgaaaa tgggcaccgt cctgacgagc ttctacggca aacgacgacc
121	ggaaaggagg tcgttcgaaa tctgcatgga gacgcggcag atactgtgga ggcgacagac
181	tgggaggaca gacggagcag ttaaaattcg tgagataaaa gagattcgtc cggtaagaa
241	ctcacgagac ttcgagaggt ggccggatga agccaagaag tatgatacct cgctctgtct
301	tgtcatatgc tacggtgccc agttcagact caagagcttg tccgtcgttg cggcaatgc
361	cgatgaacga cacaagtgga tcgtcggcct caactggcta gtggaagacc ataaaatctc
421	aagttacca agcagactag aatggtggtt acgacgggag ttctacgcca tggggaaaac
481	aaagaatgat acggtgtcac ttagggacat gaagtcattc atgcatatcg tcaacctgaa

Figure 5: the ph domain sequence from origin sequence ranging from nucleic acids( 26 – 433) with primers are highlighted in yellow

[https://www.ncbi.nlm.nih.gov/nuccore/40365362?log\\$=activity](https://www.ncbi.nlm.nih.gov/nuccore/40365362?log$=activity)

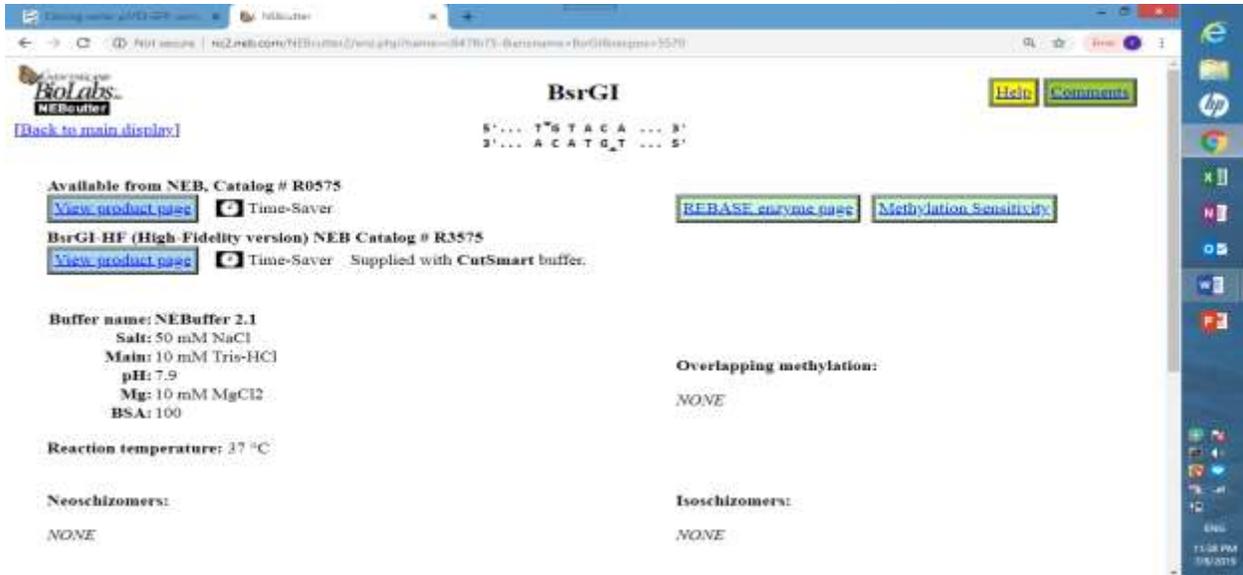


Figure 6: Picture of initial NEbcutter shows the detail of BsrGI restriction enzyme.

<http://nc2.neb.com/NEBcutter2/enz.php?name=c847fb75-&enzname=BsrGI&recpos=5570>

Next, we will use the restriction enzyme BsrGI that cut out the primers match the sites chosen for the ph domain, so GFP – PH fusion protein can be made.

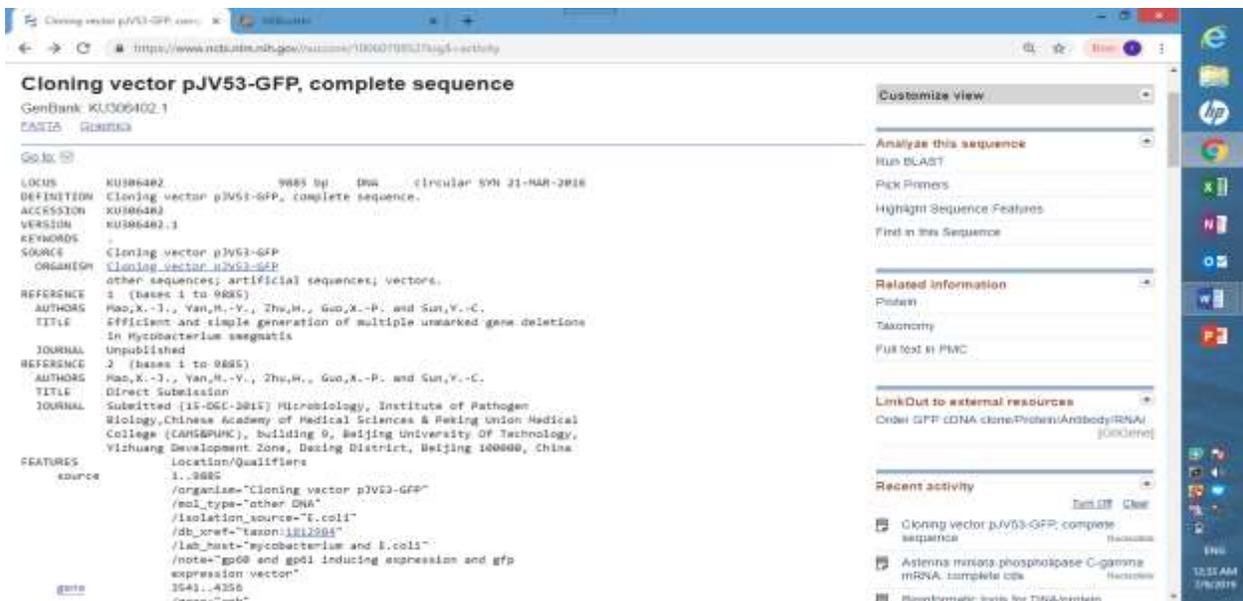


Figure 7 : pJV53-GFP cloning vector DNA sequence found in NCBI.



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AGGGCGACGCCACC TACGGCAAGCTGACCCCTGAAGTTCATCTGCACCACC
GGTAAGCTGCCGGTCCC GTGGCCGACCCCTGGTCAACCACCCTGACCTACGG
CGTCCAGTGCTTCTCCCGCTACCCGGACCACATGAAGCGCCACGACTTCT
TCAAGTCCGCCATGCCGGAGGGTTACGTCCAGGAGCGCACCATCTCCTTC
AAGGACGACGGTAAC TACAAGACGCGTGCAGGTTCAAGTTCGAGGGCGA
CACCCCTGGTCAACC GCATCGAGCTGAAGGGCATCGACTTCAAGGAGGACG
GTAACATCCTGGGCCACAAGCTGGAGTACAAC TACAAC TCCCACAACGTC
TACATCACCGGACAAGCAGAAGACGGCATCAAGGCCAACCTCAAGAC
CCGCCACAACATCGAGGACGGTGGCGTCCAGCTAGCCGACC ACTACCAGC
AGAACACCCCGATCCGGCGACGGCCCGGTCCTGCTGCCGGACAACCACTAC
CTGTCCACCCAGTCCGCCCTGTCCAAGGACCCGAACGAGAAGCGCGACCA
CATGGTCTTGCTGGAGTTCGTCAACCGCCGCGGCATCACCCACGGGCATGG
ACGAGCTGTACACAGCGTCAACCAAGATGCTGAAATGTTGGGCACCGTCTTG
ACGGCTTCTACGGCAAACGACGACCGGAAAGGAGGTTCGTTGAAATCTG
CATGGAGACGCGGCAGATAC TGTGGAGGCGACAGACTGGGCGGACAGACG
GAGCAGTTAAAATTCGTGAGATAAAGAGATTCGTCCCGGTAAGAATCA
CGAGACTCGAGAGGTGGCCGGATGAAGCTAAGAAGTATGATACCTCGCT
CTGTCTTGTATATGCTACGGTGCAGGTTACAGACTCAAGACTTGTCCG
TCGTTGCCGGCAATGCCGATGAACGACACAAGTGGATCGTCCGGCCTCAAC
TGGCTAGTGGAAATGTACAAGTAGATTTATCACCAGCCCCTCATCGTACTA

```

Figure 8: Combined PH domain with pjv53-GFP cloning vector and restriction enzyme highlight green . This sequence was copied and pasted into ORF Finder.  
<https://www.ncbi.nlm.nih.gov/orffinder/>



Figure 9:BLAST results showing successful production of GFP-PH fusion protein.  
<https://blast.ncbi.nlm.nih.gov/Blast.cgi>

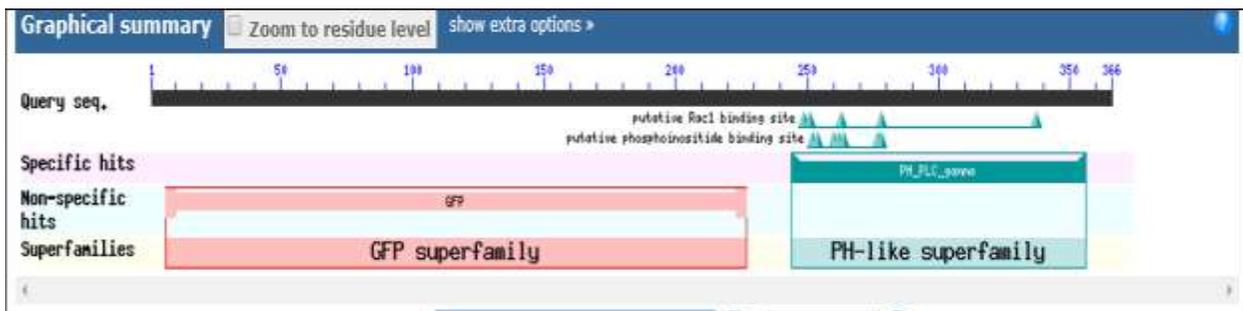


Figure 10: Zoomed-in version of successfully made GFP-PH fusion protein  
<https://blast.ncbi.nlm.nih.gov/Blast.cgi>



The egg's  $Ca^{2+}$  levels grow in response to the sperm during fertilization, which is crucial in getting the egg to start developing at the very least,  $Ca^{2+}$  when  $IP_3$  levels rise, it is released from the endoplasmic reticulum is responsible for the rise in  $Ca^{2+}$  in echinoderm and vertebrate eggs [20] [21]. However, it has not been determined how  $IP_3$  is generated during fertilization [22] [23].

A phospholipase C enzyme is responsible for this (PLC),  $IP_3$  is generated from  $PIP_2$ . The enzymes in this group contains  $\delta$ ,  $\gamma$ , and  $\beta$  isoforms. PLC  $\beta$  is activated by G proteins, whereas tyrosine kinases activate PLC [24]. Despite the fact that all three an increase in  $Ca^{2+}$  can trigger PLC isoforms the control of PLC  $\delta$  remains a mystery, even if the enzymatic activity of all three PLC isoforms may be stimulated by an increase in  $Ca^{2+}$  [25] [26] [27]. One of these phospholipase C isoforms is activated most likely leads in the production of  $IP_3$  during fertilization.

Eggs contain PLC  $\gamma$  and PLC  $\beta$  pathway proteins. For example, expression of PLC pathway/ G protein -dependent receptors such as the serotonin 2c or muscarinic m1 receptors allows for  $Ca^{2+}$  release in eggs when the appropriate antagonists are used [28] [29]. This implies the presence of functional PLC $\beta$  and related G proteins. Exogenous tyrosine kinase/PLC  $\gamma$  receptors, such as those for PDGF or EGF, can be expressed in frog and starfish eggs to allow  $Ca^{2+}$  release as a result of exposure to these agonists.  $Ca^{2+}$  release is not caused by receptors with a single point mutation that don't activate PLC $\gamma$ . A functional PLC is evident from these data. These studies have not been done on mammalian eggs, but immunoblotting has shown the presence of PLC  $\gamma$ .

## Conclusion

Several prior research have looked into whether PLC $\gamma$  or PLC $\beta$  pathways are responsible for  $Ca^{2+}$  release during fertilization. Because of concerns about the selectivity of the pharmaceutical inhibitors used, the results of these trials have not been conclusive. To determine whether PLC $\gamma$  - or PLC $\beta$ -mediated  $Ca^{2+}$  release mechanisms are involved in fertilization, we used a recombinant PLC protein component to inject starfish eggs, and it inhibited PLC $\gamma$  activation but not PLC $\beta$  activation.

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